

This article was downloaded by:

On: 25 January 2011

Access details: *Access Details: Free Access*

Publisher *Taylor & Francis*

Informa Ltd Registered in England and Wales Registered Number: 1072954 Registered office: Mortimer House, 37-41 Mortimer Street, London W1T 3JH, UK



Journal of Liquid Chromatography & Related Technologies

Publication details, including instructions for authors and subscription information:

<http://www.informaworld.com/smpp/title~content=t713597273>

Liquid Chromatographic Determination of Oxytetracycline and Chlortetracycline Residues in Animal Tissues

S. Horii^a

^a The Tokyo Metropolitan Research Laboratory of Public Health, Tokyo, Japan

To cite this Article Horii, S.(1994) 'Liquid Chromatographic Determination of Oxytetracycline and Chlortetracycline Residues in Animal Tissues', *Journal of Liquid Chromatography & Related Technologies*, 17: 1, 213 – 221

To link to this Article: DOI: 10.1080/10826079408013446

URL: <http://dx.doi.org/10.1080/10826079408013446>

PLEASE SCROLL DOWN FOR ARTICLE

Full terms and conditions of use: <http://www.informaworld.com/terms-and-conditions-of-access.pdf>

This article may be used for research, teaching and private study purposes. Any substantial or systematic reproduction, re-distribution, re-selling, loan or sub-licensing, systematic supply or distribution in any form to anyone is expressly forbidden.

The publisher does not give any warranty express or implied or make any representation that the contents will be complete or accurate or up to date. The accuracy of any instructions, formulae and drug doses should be independently verified with primary sources. The publisher shall not be liable for any loss, actions, claims, proceedings, demand or costs or damages whatsoever or howsoever caused arising directly or indirectly in connection with or arising out of the use of this material.

LIQUID CHROMATOGRAPHIC DETERMINATION OF OXYTETRACYCLINE AND CHLORTETRA- CYCLINE RESIDUES IN ANIMAL TISSUES

SHOZO HORII

*The Tokyo Metropolitan Research Laboratory of Public Health
24-1, Hyakunincho 3-chome
Shinjuku-ku, Tokyo 169, Japan*

ABSTRACT

A simple and sensitive method has been developed for the simultaneous determination of oxytetracycline and chlortetracycline in animal tissues by liquid chromatography(LC). Tissue samples were extracted with EDTA-McIlvaine buffer(pH4.0)/methanol(3/7). The extracts were purified with Sep-pak C18 and Bond Elut SCX cartridges. Two tetracyclines were separated with a synthetic polymer based-ODS column under basic condition. They were determined by LC with programmable fluorometric detector.

INTRODUCTION

Oxytetracycline(OTC) and chlortetracycline(CTC) belong to groups of tetracycline antibiotics(TCs) that are used most frequently with respect to veterinary medicine, animal nutrition and feed additives. Therefore, monitoring of residual TCs is important from the viewpoint of veterinary food hygiene. Many methods have been developed for the determination of simultaneous or individual OTC and CTC in biological samples (1-8). These meth-

ods have been based mainly on liquid chromatography (LC) with UV(1-6) or fluorometric(7,8) detection. But they lacked sensitivity required to detect TCs residues and involved tedious manipulations. This paper describes a rapid and sensitive method for the determination of OTC and CTC in tissues by LC with programmable fluorescence detection and ODS based poly(styrene divinylbenzen) copolymer as the stationary phase. This method is suitable for routine analysis of residual OTC and CTC in animal tissues.

EXPERIMENTAL

Chemicals

OTC hydrochloride and CTC hydrochloride were kindly donated by Pfizer Pharmaceuticals Inc. (Tokyo, Japan) and Takeda Chemical Industry Ltd. (Osaka, Japan), respectively. Acetonitrile was of LC grade and all other chemicals were of analytical grade from Cica-Merck (Tokyo, Japan). Sep-pak C18 and Bond Elut SCX cartridges were purchased from Waters Association (Milford, MA, U.S.A.) and Analytichem International (Harbor City, CA, U.S.A.), respectively. LC grade water was obtained by purifying reversed osmosis water in a Milli-Q II system (Millipore, Bedford, MA, U.S.A.).

Apparatus and chromatographic conditions

An LC system consisted of two 6AD pumps, a SIL-6B auto injector, a SCL-6B system controller, a CTO-6A column oven, a RF-550A spectrofluorometer, a C-R4AX integrator (Shimadzu, Kyoto, Japan) and a KT-35 degasser (Shodex, Tokyo, Japan). All analyses were carried out using a ODP-50 5-um 250mm x 4.6mm (I.D.) column, which was purchased from Asahi chemical ind. (Tokyo, Japan). The mobile

phase was prepared by mixing 900 ml of Sorensen buffer (pH 12.0) and 100 ml of acetonitrile. The flow rate was 1 ml/min and the column temperature was maintained at 40°C. The wave length of spectrofluorometer was changed to ex. 350nm, em. 420nm from ex. 374nm, em. 508nm 7 min later, by using time programming technique.

Sample preparation procedure

Minced 10g tissue was homogenized with 30ml of 10mM Na₂EDTA-McIlvaine buffer (pH 4.0)-methanol (3:7) mixture and centrifuged twice. The supernate was combined and concentrated to ca. 5 ml under vacuum. The concentrated solution was offered onto a Sep-pak C18 cartridge column. The cartridge was rinsed with 5 ml of water and eluted with 10 ml of methanol. The eluate was offered onto the next Bond Elut SCX cartridge. The cartridge was rinsed with 10 ml of water and eluted with 10 ml of 1N HCl-methanol (2:8). The eluate was adjusted to pH 12 with 10N sodium hydroxide and injected to the LC after an hour.

RESULTS AND DISCUSSION

Variation of the emission spectra of OTC(I) and CTC(II) for different pH (9-12) solution are shown in Fig.1. The maximum fluorescence wave length for OTC is at 390nm on excitation, and 495-500nm on emission, and that for CTC is 350nm on excitation, and 405-410nm on emission, respectively. As CTC decomposes in alkaline medium to form isochlortetracycline (ISOCTC) (7), the fluorescence intensities of TCs are strongly dependent on the pH of the solution. Perhaps, the same phenomenon may happen on OTC to that on as CTC. The reduced fluorescence intensities in alkaline medium are due to the decomposition of these two TCs.

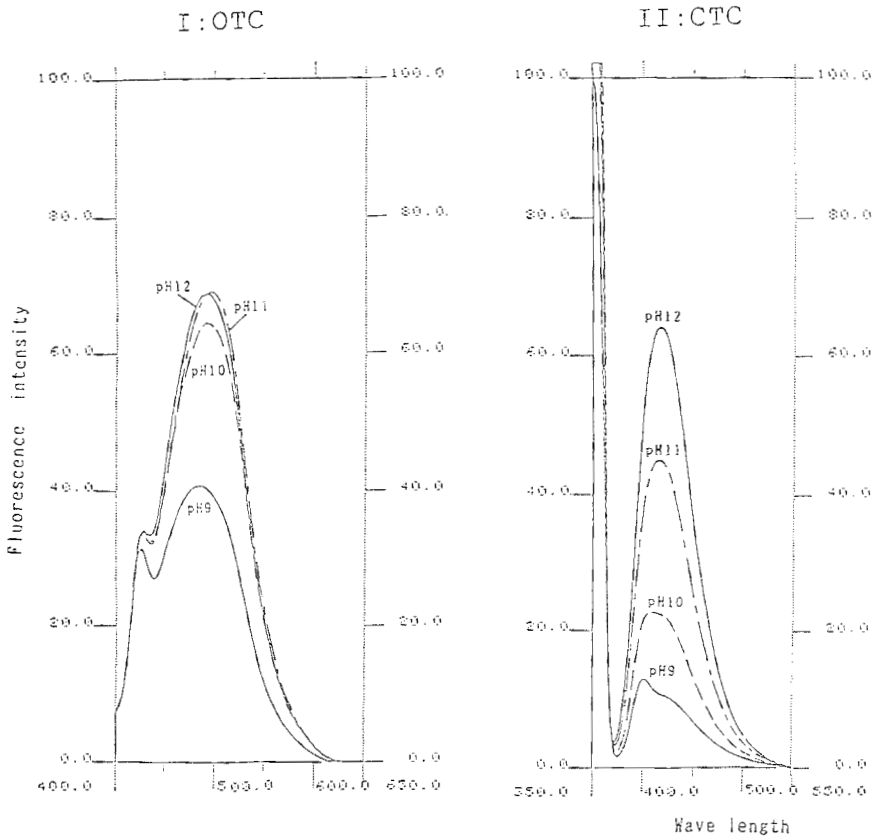


Fig. 1. Emission spectra of TCs (I:OTC,II:CTC) for different PH(9-12) solutions.

The time-dependent changes of decomposed TCs are shown in Fig.2,3. The fluorescence intensity of OTC reached the maximum value immediately and was held constant for long time. That of CTC increased slowly and reached plateau after an hour, to maintain its intensity for moreover 24 hrs. The reacted time was of one hour was necessary for the simultaneous determination of these two TCs.

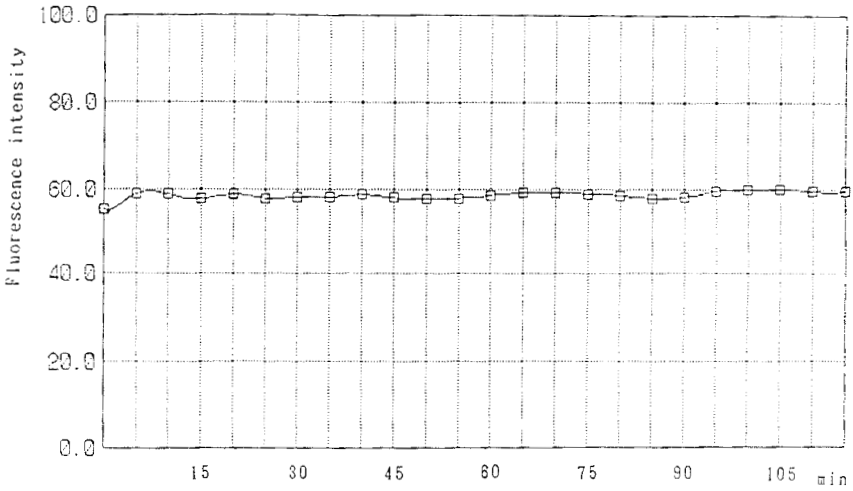


Fig.2. Fluorescence intensity versus time profile of OTC in basic condition (mobile phase).

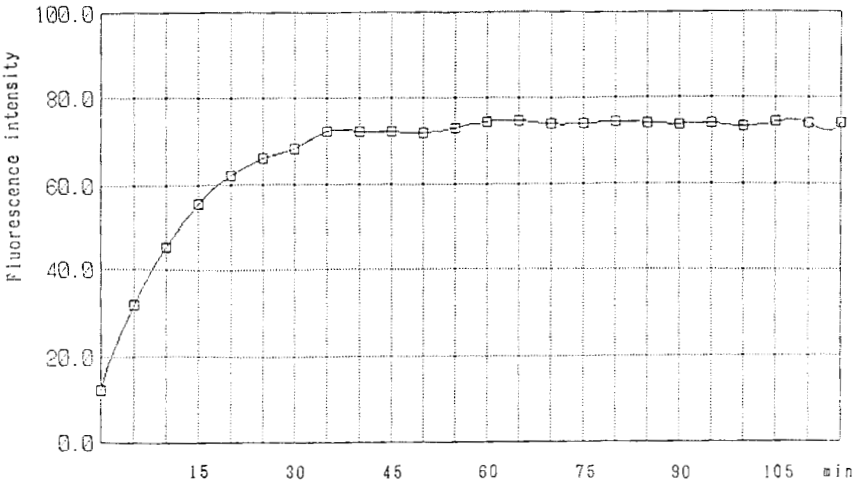


Fig.3. Fluorescence intensity versus time profile of CTC in basic condition (mobile phase).

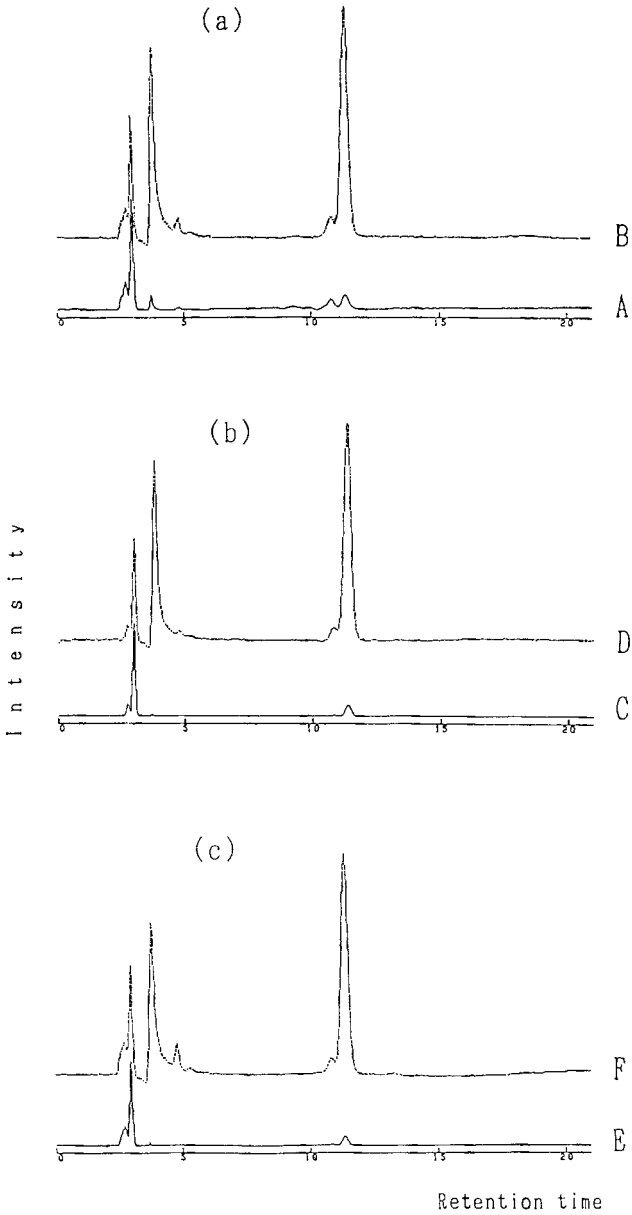


Fig. 4. Liquid chromatograms of drug free & fortified samples; A, C, E: drug free samples; B, D, F: fortified samples; (a): bovine; (b): swine; (c): chicken; LC conditions are described in literature.

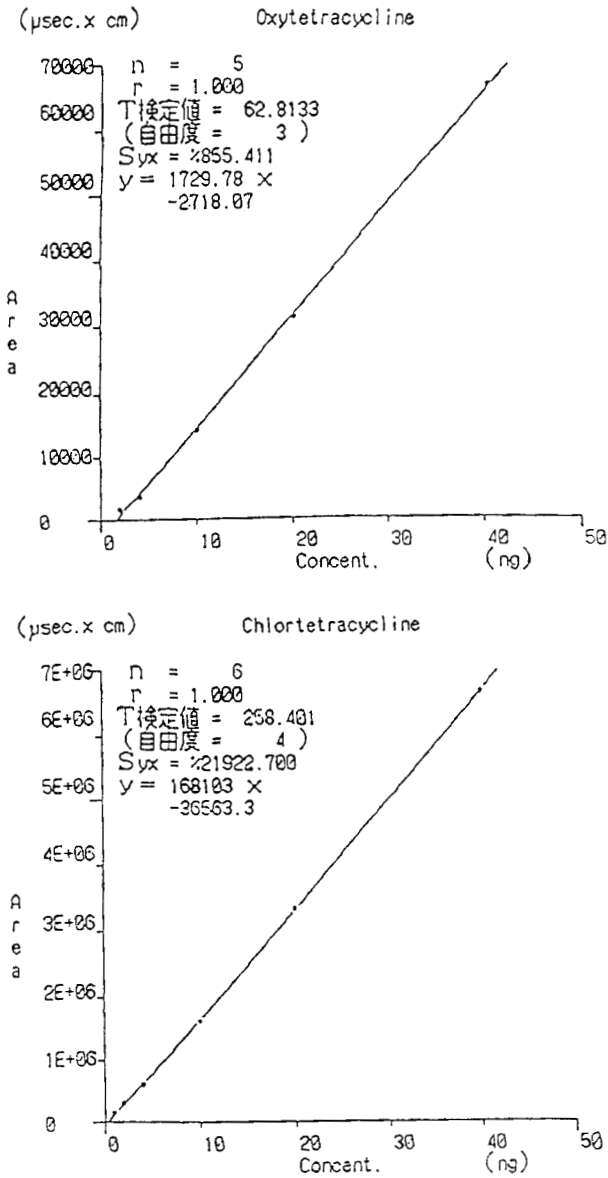


Fig. 5. Calibration curves for OTC and CTC.

Table 1 Recoveries of OTC and CTC from Bovine, Swine and Chicken.

	Added ($\mu\text{g/g}$)	Recovery \pm S. D., C. V. (%)					
		Bovine		Swine		Chicken	
OTC	0.2	79.5 \pm 5.4, 6.8	74.4 \pm 4.3, 5.8	63.4 \pm 1.6, 2.5			
	1.0	81.3 \pm 7.4, 9.1	85.6 \pm 3.9, 4.6	74.8 \pm 4.6, 6.1			
CTC	0.2	78.5 \pm 6.1, 7.8	72.3 \pm 6.3, 8.7	61.8 \pm 3.6, 5.8			
	1.0	86.5 \pm 9.7, 11.2	73.1 \pm 6.0, 8.2	70.5 \pm 4.5, 6.4			

n=5

Some large interfering peaks derived from the matrix appeared before OTC and CTC peaks in the case of using an only ODS cartridge. The second cartridge of SCX was therefore connected with the ODS cartridge to remove the interfering peaks. Although they still appeared after the use of double cartridges, this method was fairly effective in minimizing the interference with co-extracted components of the meat. Both peak height of 5ng OTC 1ng CTC were more than 4 times that of the interfering peaks. Consequently, the matrix peaks were estimated to be negligible. These results are shown in Fig.4.

Results of calibration runs for the validation range of 2-40ng (OTC) and 1-40ng(CTC), respectively seen in Fig.5, showed excellent linearity ($r=0.999$).

Two amounts(0.2 μg) of TCs were added to the drug free minced meat(10g), and their recoveries were measured by the procedure described above. The results are summarized in Table 1, which shows recovery ranges from 61.8 to 86.5% and coefficients of variation (C.V.) from 2.5 to 11.2%.

REFERENCES

1. Onji, Y., Uno, M. and Tanigawa, K., Liquid Chromatographic Determination of Tetracycline Residues in Meat and Fish, *J. Assoc. Off. Anal. Chem.*, 67, 1135, 1984.
2. Ikai, Y., Oka, H., Kawamura, N. and Yamada, M., Improvement of Chemical Analysis of Antibiotics, *J. Chromatogr.*, 411, 313, 1987.
3. Rogstad, A., Hormazabal, V. and Yndestad, M., Optimization of Solid Phase Extraction of Oxytetracycline from Fish Tissue and Its Determination by HPLC, *J. Liq. Chromatogr.*, 11, 2337, 1988.
4. Mulders, E.J. and Van De Langemaat, D., Determination of Residues of Tetracycline Antibiotics in Animal Tissues by HPLC, *J. Pharm. Biomed. Anal.*, 7, 1829, 1989.
5. Thomas, M.H., Simultaneous Determination of Oxytetracycline, Tetracycline, and Chlortetracycline in Milk by Liquid Chromatography, *J. Assoc. Off. Anal. Chem.*, 72, 564, 1989.
6. Farrington, W.H.H., Tarbin, J., Bygrave, J. and Shearer, G., Analysis of Trace Residues of Tetracyclines in Animal Tissues and Fluids Using Metal Chelate Affinity Chromatography/HPLC, *Food Addit. Contam.*, 8, 55, 1991.
7. Khan, N.H., Roets, E., Hoogmartens, J. and Vanderhaeghe, H., Quantitative Analysis of Chlortetracycline and Related Substances by HPLC, *J. Pharm. Biomed. Anal.*, 7, 339, 1989.
8. Blanchflower, W.J., McCracken, R.J. and Rice, D.A., Determination of Chlortetracycline Residues in Tissues Using HPLC With Fluorescence Detection, *Analyst*, 114, 421, 1989.

Received: June 6, 1993

Accepted: June 10, 1993